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## Literature Review - Pigments in Plant Health

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Applicability of Pigment Compounds for Improving Plant Health

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Literature Review

### Abstract

The solar radiation received at the Earth's surface helps power the photochemical processes in photosynthesis. However, under sunlight, photosynthetic organisms are unavoidably exposed to potentially harmful levels of ultraviolet and visible light radiation. Consequently, plants have developed many mechanisms for the avoidance and dissipation of excess light energy. The growth of crops under increased solar irradiance during sub- and supra-optimal temperature conditions is an obstacle for increasing seasonal crop production in the transition zone. New methods for reducing the photoinhibitory effects of solar irradiation have been observed in the field of turfgrass. Through the application of pigment compounds to the surface of plant leaves, it may be possible to reduce levels of solar incident light exposure to the plant, during temperatures where the fixation of CO<sub>2</sub> is a limiting factor in Photosystem II repair. This proposal will explore the applicability of various commercial and naturally derived pigment compounds for improving the photosynthetic capacity of creeping bentgrass plants under supraoptimal light and sub- and supra-optimal temperature stress.

### **Creeping Bentgrass**

Creeping bentgrass (*Agrostis palustris* ssp. *stolonifera* L.) is a fine textured C3 turfgrass used on golf course greens due to its ability to tolerate low-mowing heights (Warnke, 2003). The aesthetic and performance properties of creeping bentgrass has led to its use in supraoptimal temperature climates, where its health will decline in summer months (Carrow, 1996). For cool-season turfgrasses, ambient air temperatures above 24 C and soil temperatures above 18 C are considered to be supraoptimal for shoot and root growth, respectively (Beard, 1973). Hot and humid conditions in the southeast can produce temperatures above these optimal levels, resulting in a range of symptoms characterized as Summer Bentgrass Decline (SBD) (Carrow, 1996). High air and soil temperatures lead to an increased respiration rate in bentgrass roots, and subsequent dieback upon carbohydrate depletion (Xu and Huang, 2000; Xu & Huang, 2001). In particular, supraoptimal soil temperatures have been shown to be most detrimental to creeping bentgrass root and shoot growth and nutrient uptake (Xu & Huang, 2000). Heat can also disrupt the plant's cellular membranes, severely affecting cellular function (Larkindale & Huang, 2004).

Evaporation and transpiration release heat energy through latent heat of vaporization, which describes the transfer of energy in the state change of water from a liquid to a gas form. Conversely, the same amount of energy is released when water condenses from vapor to liquid form. In other words, evaporation will cool a surface, while condensation will warm it (Tarara, 2000). Differences between water vapor concentration in and outside the plant will determine rates of transpiration. Under conditions of increased humidity, transpirational cooling will decrease, due to reductions in the water vapor concentration gradient between the plant and the air (Rawson, Begg, & Woodward, 1977). However, an increased surface temperature will cause an exponential increase in water vapor concentration at the surface, raising this gradient, which leads to greater transpirational flow of water vapor from the plant.

Consequently, very small changes in temperature can trigger large fluxes in the rate of transpiration (Gates, 1965; Pallardy & Kozlowski, 2008).

Radiation is the largest contributor to increasing soil temperatures (Huang, 2002; Larkindale & Huang, 2004). The root zone of USGA (United States Golf Association) certified golf course greens incorporate high amounts of sand near the surface of the soil (Moore, 2004). Among various soil-types, Abu-Hamdeh & Reeder (2000) observed the highest thermal conductivity in sandy soils. The temperatures of the various soils were positively correlated with bulk density. Research has shown that the addition of organic matter will decrease bulk density (Hummel, 1993). Creeping bentgrass cultivars utilized in golf course greens exhibit high rates of organic matter accumulation at the soil surface (Carrow, 2004; Hudson & Shane, 1994). This accumulation of organic matter will reduce bulk density, and thus thermal conductivity. However, organic matter can also increase water retention, which dramatically increases thermal conductivity with increases in temperature (Abu-Hamdeh & Reeder, 2000; Campbell, Jungbauer, Bidlake, & Hungerford, 1994). Bare portions of the turf will also expose surface organic matter, which will absorb higher amounts of incident light energy due to its darker color (Bristow & Horton, 1996; Loughrin & Kasperbauer, 2003). Heat will either move upward to the surface of soil, or downward from warmer to cooler layers (Tarara, 2000).

Increases in leaf lipid saturation levels have been observed following heat treatment to creeping bentgrass plants (Larkindale & Huang, 2004). The thylakoid membrane contains a large proportion of non-bilayer forming lipids, which are thought to be required for the stabilization of contained Photosystem-II molecules (Thomas et al., 1986). Increases in lipid saturation, within chloroplast structures, allow for a higher tolerance of temperatures, reducing phase-separation of non-bilayer

forming lipids from the chloroplast membrane (Gounaris et al., 1983). However, no changes in lipid composition concentration were observed in the roots of creeping bentgrass (Larkindale & Huang, 2004). Incidences of heat damage have been shown to be light-mediated, supporting the idea that any subsequent damage could occur as a result of light induced damage to the photosynthetic apparatus (Larkindale & Knight, 2002). This idea is supported in creeping bentgrass species, where photosynthetic acclimation has been shown to be essential for increases in tolerance to severe heat stress, due to maintenance of light-harvesting capacity and carbon fixation throughout the heat stress period (Liu & Huang, 2008). The specific mechanisms relating to light induced inhibition of the photosynthetic apparatus will be further discussed later.

### **Light and Plant Pigments**

Visible light (400-700 nm) accounts for 43% of the energy in the global solar irradiance spectrum (300-2500 nm) for North America. The remainder of this energy arrives as 52% near-infrared (NIR) (700-2500) and 5% ultraviolet (UV) (300-400 nm) (Levinson et al., 2005*b*; American Society for Testing and Materials [ASTM], 2003). NIR wavelengths are responsible for much of the heating within leaves (Forbes & Watson, 1992). Consequently, plant leaves will effectively scatter and reflect 70% of incident perpendicular infrared radiation; reducing heat-buildup in the plant and the soil (Knipling, 1970; Atwell, Kriedemann, & Turnbull, 1999; Larcher, 2003). Plants exhibiting bicolouration (abaxial surface a lighter shade of green than adaxial), a thicker cuticle, and a higher portion of mesophyll surface area exposed to intercellular air spaces per unit leaf surface area exhibited predictably higher NIR reflectance values from the adaxial leaf surface. However, in plants with lower pigment content, less incident UV-visible light would be absorbed and less NIR light would be reflected, allowing greater amounts of heat energy to build in the soil, as well as be reradiated as heat. Kopsell, Lefsrud, Sorochan, & McElroy (2010) reported that, among

heat-tolerant cultivars of *Poa pratensis*, those highest in pigment content were the least heat tolerant. However, also mentioned is the possibility that drought was an influencing factor in decreasing pigmentation across cultivars, which would account for a decrease in transpirational cooling and CO<sub>2</sub> fixation. Dry plant leaves will also show diminished reflectance in the NIR, which would increase NIR absorbance by the soil (Hawley, 1971).

Light consists of both wave and particulate properties. While light is propagated in wave form, its interaction with matter is more particulate (Prasad, 1997). Planck's law (Planck, 1989) describes how light exists in quanta, or bundles of energy. This theory was further developed by Einstein (1905), who went on to discover that radiation processes involve the emission or absorption of light quanta, or "photons" (Bohr, 1949). The energy of a photon is determined by its wavelength, the distance between repeating waves, and frequency, the number of light wave repetitions per unit time. Their relationship is considered directly proportional, in that any fraction of photon wavelength equals its reciprocal in energy output, and is a function of frequency (Prasad, 1997). This is not to be confused with the intensity of light, which depends on how many photons of energy are being emitted per unit time (McDonald, 2003).

Two basic principles of light that govern the absorption properties of plant pigments are the Grotthus-Draper and Stark-Einstein laws. The Grotthus-Draper law states that photochemical processes can only occur with absorbed radiation, and the Stark-Einstein law states that each absorbed photon can only affect one molecule (Diffey, 2002). When plant pigment molecules in an unexcited ground state absorb photons of a compatible wavelength, a valance electron is quickly raised to an excited state. After returning to a ground state, the absorbed energy is released in the form of thermal dissipation, fluorescence, phosphorescence, or inductive resonance (McDonald, 2003). However, long-term exposure

of plants to supraoptimal levels of light can result in the photodestruction of photosynthetic pigments. This chemical reaction is considered oxygen- and light-dependant, and is defined as photooxidation (Powles, 1984). Light toxicity occurs when high flux light converts the target pigment molecules into an excited state, first, an initial short-lived singlet state, followed by molecular rearrangement into a longer-lived triplet state. During this time, there is an increased chance for chemical reactions with surrounding molecules. In the formation of damaging species, triplet energy is transferred to molecular oxygen, forming singlet oxygen. This reactive oxygen species exists as a free radical, capable of oxidizing and bleaching plant pigments to an irreversible degree under extreme conditions (Larson, 1988). However, plants have evolved multiple protective mechanisms for the effective removal of this excess light energy.

Photosynthetic systems are composed of a network of principal and accessory pigments (Duysens & Ames, 1962). While principal pigments are directly involved in the chemical conversion of energy, accessory pigments act as sinks for conducting different wavelengths, transferring the energy to the primary through inductive resonance. Together, these pigments form a network of overlapping absorption bands, improving the efficiency at which plants harvest light in the action spectrum of photosynthesis (Smith & French, 1963; McCree, 1971). However, accessory pigments also play an important role in photoprotection.

Functioning as an accessory pigment, carotenoids of the xanthophyll cycle are considered essential to the dissipation of energy under conditions of excess light, through the interception and removal of excess excitation energy prior to its entrance into the electron transport chain (Demmig-Adams & Adams III, 1996). Once the absorption of light exceeds a plant's capacity for CO<sub>2</sub> fixation, photosynthetic electron transport generates a decrease in lumen pH. This activates the conversion of xanthophylls, removing oxygen from violaxanthin

to form antheraxanthin, then zeaxanthin, which dissipates the excess energy as heat (Muller, 2001).

Flavanoids, another group of accessory pigments, make up one of the largest known groups of phenolic compounds within plants, with over 9000 assessed from plant tissue as reported by Williams & Grayer (2004). Responsible for the many bright blue, red, and purple colors throughout nature, anthocyanins are the most widespread of the pigmented flavonoids (McDonald, 2003). The prospective roles of anthocyanin in plants are numerous, and have been contemplated by scientists for well over a century (Gould, 2010). As stated by Wheldale (1916), in one of the earliest reviews of plant anthocyanin function, “It is difficult to find a hypothesis which fits all cases of anthocyanin distribution without reduction to absurdity.” However, in recent years there have been many significant advances in understanding the roles of anthocyanin pigments in plants. Related to its light-attenuation properties, anthocyanin accumulation has been linked to photoprotection of chlorophyll during drought and cold stress (E. Taulavuori, Tahkokorpi, Laine, & K. Taulavuori 2010, Gould, Dudle, & Neufeld, 2010), improved recovery from mechanical injury (Gould, McKelvie, & Markham, 2002), enhanced nutrient retrieval from senescing leaves (Hoch, Singaas, & McCown, 2003), and delayed senescence in CO<sub>2</sub> rich environments (Tallis, Lin, Rogers, Zhang, Street, Miglietta, Karnosky, De Angelis, Calfapietra, & Taylor, 2010). Independent of its light attenuation properties, anthocyanin has also been ascribed to many biotic-dependant roles, including: microbial defence responses (Kangatharalingam, Pierce, Bayles, & Essenberg, 2002; Hipskind, Wood, & Nicholson, 1996); herbivory avoidance (Karageorgou & Manetas, 2006); and pollination ecology (Harborne & Smith, 1978). Due to the diverse range of inducing factors associated with anthocyanin biosynthesis, correlating the transient

accumulation of anthocyanin to any one function is inherently difficult. Consequently, knowledge of the localization and spectroscopic properties *in vivo* of all the pigment pools is essential for ecophysiological studies and the quantitative description of anthocyanin function (Gould et al., 2002). While its distribution in plants differs considerably across species, anthocyanin will generally localize within cell vacuoles, in or just below the adaxial epidermis, effectively providing light-protection to subjacent chloroplasts (Merzlyak, Melo, & Naqvi, 2008). Abaxial accumulation in leaves has been observed, but is also considered a photoprotective adaptation of light-sensitive plants whose leaf orientation and substrate albedo may vary throughout developmental stages of the plant (Hughes & Smith, 2007).

Anthocyanin's ability to protect plant tissue during events like wounding, drought, and pathogen infection can also be considered an indirect effect of light screening, due to an avoidance of light-dependant oxidation of tissue through reductions in excitation pressure during periods of increased tissue vulnerability (Steyn, Wand, Holcroft, & Jacobs, 2002). In light of this observed reduction in the accumulation of reactive oxygen species, there has been an inescapable interest in the potential role of flavanoids as antioxidants *in vivo*. A study by Rice-Evans, Miller, & Paganga (1997), demonstrated the *in vitro* antioxidant capacity of plant-derived flavonoids to be several times more effective than both ascorbate (vitamin C) and  $\alpha$ -tocopherol (vitamin E), two functional antioxidants in plants. This would suggest potential antioxidant benefits associated with flavonoid presence *in vivo*. However, the chemical diversity and complex biochemistry of flavonoids *in vivo* have prevented adequate correlations between flavonoid accumulation, flavonoid oxidation products, and oxidative stress. Consequently, the putative function of flavonoids as antioxidants is still a matter of debate (Hernández, Alegre, Van Breusegem, & Munné-Bosch, 2009).

### **New Models of Photoinhibition**

Photosynthesis requires the interaction between two separate, but equally complex photosystems. Photosystem I (PSI) and Photosystem II (PSII) have designations of P700 and P680 respectively, named for the absorption maximum of their chlorophyll *a* molecules. Each of the photosystems contains subtle differences in protein associations, which accounts for their different absorption properties (Anderson & Andersson, 1988). Chlorophyll *a* molecules only absorb a small portion of light for use in photosynthesis. In order to better utilize the reaction centres in photosynthesis, each photosystem utilizes antenna complexes composed of several hundred pigment molecules. These light-harvesting complexes help to extend the absorption spectrum, using resonance transfer to designate the flow of excitation energy to the reaction centres (Glazer, 1989; Hunter, van Grondelle, & Olsen 1989; Zuber, 1986).

The absorption of excess light energy has the potential to damage photosynthetic machinery, beginning with PSII. Photoinactivation of PSII is hypothesized to occur by two separate mechanisms, acceptor-side and donor-side, which both result in the inhibition of electron transfer in PSII, and subsequent degradation of the D1 protein (Wei, Cady, Brudvig, & Hou, 2011). PSII reaction centre D1 proteins, which exhibit the highest turnover rates in the thylakoid membrane, are the main target of oxidation during photodamage (Sunby, McCaffery, & Anderson, 1993). In the acceptor-side mechanism hypothesis, photoinhibition begins with the reduction of the plastoquinone pool under intense light, causing a lack of oxidized plastoquinone to bind to the  $Q_B$  site on the D1 protein. Because  $Q_A^-$  is unable to transfer an electron to  $Q_B$ , it becomes doubly reduced to  $Q_A^{2-}$  during the second turnover of the reaction centre. In order to become stable,  $Q_A^{2-}$  will become protonated, forming  $Q_AH_2$ , which is then released from the  $Q_A$  binding site on the D1 protein. The newly unoccupied  $Q_A$  site leads to the formation of the primary radical pair  $P680^+Pheo^-$ , and through recombination, generates triplet state P680. This reacts with oxygen to form singlet oxygen, a

reactive oxygen species responsible for D1 protein degradation (Wei et al., 2011; Tyystjärvi, 2008; Anderson, Park, & Chow, 1998). In the donor-side mechanism hypothesis, highly reactive  $P680^+$  is formed due to a lack of electron donation while under the influence of light.  $P680^+$  will oxidize surrounding chlorophyll and carotenoid molecules, and lead to degradation the D1 protein (Wei et al., 2011).

Under experimental light conditions, photodamage to PSII was found to be greatest in regions of UV and yellow light exposure in *Arabidopsis thaliana* (Takahashi, Milward, Yamori, Evans, Hillier, & Badger, 2010). Damage to PSII by high energy light can best be explained by a new two-step model developed by Ohnishi, Allakhverdiev, Takahashi, Higashi, Watanabe, Nishiyama, & Murata, (2005) and further explained by Tyystjärvi (2008). This theoretical model states that photoinhibition begins with the reduction of the Mn cluster in the oxygen-evolving complex by UV, blue, and green light, but not red (Wei et al., 2011). Following Mn inactivation, PSII becomes sensitive to light at 680 nm, experiencing inactivation from direct red and blue light exposure to its photosynthetic pigments in the donor-side mechanism (Ohnishi et al., 2005). Damage to the oxygen-evolving complex will increase potential for PSII damage due to reductions in electron donation from the oxygen evolving complex to PSII undergoing oxidation (Hakala, Tuominen, Keränen, T. Tyystjärvi, E. Tyystjärvi, 2005). Takahashi et al. (2010) attempts to explain the adverse effects of yellow light, attributing its photoinhibitory effects to the manganese light-sensitizer mechanism. While yellow light contains less excitation energy than ultraviolet (UV) and blue light, it is much more abundant in the solar spectrum (Takahashi & Badger, 2010). More of this light is able to penetrate plant tissue, due to its lack of absorption by anthocyanin (primarily blue and green light absorption), chlorophyll (primarily blue and red light absorption) and carotenoid (primarily blue and green light absorption) pigments (Takahashi et al., 2010; Solovchenko & Merzlyak, 2008). The PSII photodamage spectrum is very different from the absorption

spectra of these pigments, but is closely correlated with that of manganese compounds (Wei et al., 2011). Because collimated light is scattered within leaf tissue, the efficiency of its absorption will increase with depth in the mesophyll (Vogelman, Nishio, & Smith, 1996). This allows non-photosynthetic yellow and green light to penetrate more deeply into the leaf, and trigger excitation in shade adapted chloroplasts of the lower mesophyll (Nishio, 2000). The increased presence of refracted yellow-green light in the mesophyll may have a greater influence on Mn excitation in the absence of high energy UV light. This suggests that the mechanism of light diffusion, meant to increase the absorption of light by pigment networks, could also contribute to the indirect photodamage of PSII during high visible light irradiance (Takahasi et al., 2010). However, a recent study revealed that visible light had little impact on the production of high valent species of Mn in the oxygen evolving complex, while UV light did (Wei et al., 2011). Consequently, UV light inhibition of the oxygen evolving complex in PSII is also much faster and thus more damaging than that of visible light (Tyystjärvi, 2008). Wei et al. (2011) uses this to support the idea that excess visible light induced photodamage occurs directly to PSII, without inhibiting the Mn oxygen-evolving complex. This supports the theory that, while donor-side photoinhibition has often been observed after chemical inactivation of the oxygen evolving complex by UV light, there is still potential for visible light to trigger this mechanism in the absence of UV and blue light, because the OEC will sometimes fail to reduce highly reactive  $P680^+$  species (Anderson et al., 1998; Wei et al., 2011). The absorption peak of anthocyanin in the visible light region (450-550 nm), suggests that it may provide photoprotection in this mechanism as well (Solovchenko & Merzlyak, 2008). Overall, the close correlation between this photodamage spectrum and the anthocyanin absorbance spectrum supports the

hypothesis that adaxial localized phenolic compounds are meant to act as filters for high energy light (Takahashi & Badger, 2010).

### **PSII Repair Cycle**

Upon photodamage to PSII, the plant begins to replace damaged PSII proteins in a process known as the PSII repair cycle (Aro, Suorsa, Rokka, Allahverdiyeva, Paakkarinen, Saleem, Battchikova, & Rintamäki, 2004). In order to repair the damaged PSII complex, the photodamaged D1 protein is rapidly degraded, *de novo* synthesized, and incorporated back into PSII (van Wijk, Roobol-Boza, Kettunen, Anderson, & Aros, 1997; Nishiyama, Yamamoto, Allakhverdiev, Inaba, Yokota, & Murata, 2001). Environmental stresses can inhibit *de novo* synthesis of the D1 protein, and consequently limit the rate and extent of PSII repair (Allakhverdiev & Murata, 2004). In forming a common mechanism for this inhibitory response, the role of reactive oxygen species seems likely. The fixation of CO<sub>2</sub> is sensitive to a wide array of environmental stresses: including light (Sun, Nishio, & Vogelmann, 1996), temperature, drought (Cornic & Ghashghaie, 1991) and salt (Yeo, Caporn, & Flowers, 1985). Limitation of CO<sub>2</sub> fixation decreases NADPH use efficiency, subsequently reducing NADP<sup>+</sup>, a major acceptor of electrons in PSI. This accelerates the rate of electron transport to molecular oxygen, forming superoxide anions, which lead to formation of H<sub>2</sub>O<sub>2</sub> by superoxide dismutase in PSI (Takahashi & Murata, 2008). The reaction for scavenging of H<sub>2</sub>O<sub>2</sub> is the photoreduction of dioxygen to water in PSI by the electrons derived from water in PSII. The increased production of H<sub>2</sub>O<sub>2</sub> can exceed the rate at which it can be scavenged in the water-water cycle (Takahashi & Murata, 2008; Asada, 1999; DeRose, Mukerji, Latimer, Yachandra, Sauer, & Klein, 1994; Barber, 2008; Song, Liu, Wang, Li, & Liu, 2006). Unscavenged H<sub>2</sub>O<sub>2</sub> inhibits the repair of PSII through blocking the synthesis of a D1 precursor (Apel & Heribert, 2004; Nishiyama et al., 2001). Though these reactions have no

effect on the rate of photodamage to PSII, with repair inhibited, photoinhibition is accelerated due to on-going damage incurred from light exposure (Takahashi & Murata, 2005).

Adaxial localized screening pigments increase the reflectance of red light, the absorbance of blue light, and the attenuation of green light. In an effort to better understand the role of adaxially localized pigment compounds in preventing photoinhibition in the lower mesophyll, Hormaetxe, Becerril, Fleck, Pintó, & Garcia-Plazaola (2005) tested the filtration qualities of variously colored cultivars of *Buxus sempervirens* when under photoinhibitory conditions. Adaxial sections of green, brown, orange yellow, and red colored cultivars were positioned *in loco* of adaxial removed sections of green shade leaves. Green adaxial sections demonstrated the highest levels of absorptance, and consequently, the lowest levels of photoinhibition in the lower mesophyll. These results are most likely due to the increased light absorption efficiency and photostability of high chlorophyll content leaves, whereas the accumulation of light filtration pigments is usually associated with lower levels of chlorophyll, and thus lower light use efficiency (Close & Beadle, 2003). Assuming this higher light protection by chlorophyll in the adaxial section of the leaves of this species, researchers should begin to ask why certain plants didn't evolve specialized green pigments for the more effective filtration of excess light (Hormaetxe et al., 2005).

### **Commercial Pigments**

Over recent years there has been an increasing interest in the utilization of “stress guard” pigments in the field of turfgrass science. In 2004, a patent was filed for the commercial use of Pigment Green 7, a polychlorinated form of copper phthalocyanine, on turfgrass plants. Under supraoptimal heat stress, creeping bentgrass plants treated with copper

phthalocyanine showed increases in quality, chlorophyll content, carotenoid content, and photochemical efficiency when compared to controls.

Copper phthalocyanine; specifically Pigment Green 7 (chlorinated copper phthalocyanine), has a long history of application across many industries. It has mainly been used in outdoor paints, due to its increased dispersibility, light fastness, heat stability and durability (Kadish, Smith, & Guillard, 2003; Tracton, 2006). More recently, it has begun to see use as a photosensitizer in dye-sensitized organic solar cells (Huang, Yu, Lin, & Jiang, 2009; Chu, Shrotriya, Li, & Yang, 2006; Tripathi, Datta, Samal, Awasthi, & Kumar, 2008). This use can be attributed to the structural and spectroscopic similarities of copper phthalocyanine to plant chlorophyll compounds (Ludwig, Strohmaier, Peterson, Gompf, & Eisenmenger, 1994; Karan, Basak, & Mallik, 2007; Bohn & Walczyk, 2004). Naturally, this has resulted in many experiments evaluating their efficiency in harvesting solar energy (Frag, 2007).

Copper phthalocyanine (phthalocyanine blue) and chlorinated copper phthalocyanine (phthalocyanine green) are pigments known for their excellent light and heat stability, allowing them to maintain structure under outdoor conditions. In general, all copper phthalocyanines are considered weakly scattering pigments, with strong absorption in the red to near-infrared (NIR) portions between 500 to 700 nm, the UV region between 300 and 400 nm and equally strong fluorescence in the UV-blue between 350 to 500 nm. Its UV-blue absorption and fluorescence spectrum strongly overlap the emission band of UV light capable of high photoinhibition efficiency in the excitation of manganese in the oxygen evolving complex between 300 and 500 nm (Levinson, Berdahl, & Akbart, 2005*a*; Levinson, Berdahl, & Akbari, 2005*b*; Bigger & Delatycki, 1989; Saron, Zulli, Giordano, & Felisberti, 2006; Hakala et al., 2005). Plants are only able to reflect very small amounts of UV radiation (~3%) (Larcher, 2003). Copper phthalocyanine's ability to attenuate extremely high levels of

incident high energy light with minimal scattering, may allow it to effectively reduce levels of bentgrass photoinhibition. By adjusting the density at which the pigment is applied via a carrier (i.e. – water), the level of incident light attenuation can be adjusted to meet the high-light avoidance needs of the plant. The largest dip in light attenuation by copper phthalocyanine occurs at 550 nm, allowing it to closely mimic the action spectrum of photosynthesis, and have a green color (McDonald, 2003; Saron et al., 2006).

Pigments more capable of reflecting NIR light have been shown to be much cooler in sunlight than those that absorb NIR. This has been shown to have particular importance in roofing, where the use of roofs with NIR reflecting pigments has been shown to significantly reduce building heat gain over that of roofs utilizing NIR absorbing pigments (Levinson et al., 2005*a*; Levinson et al., 2005*b*). This has led to the identification of dark colored pigments that have the ability to reflect infrared heat-building rays to the same degree that a white roof would (Miller, Akbari, & Levinson, 2004).

In the Norton et al. (2006) patent for utilizing copper phthalocyanine to improve turfgrass quality, there was specific interest on the increased efficacy of phthalocyanine green pigments over that of phthalocyanine blue. Spectroscopically, the largest difference between phthalocyanine blue and phthalocyanine green is that phthalocyanine green absorbs more strongly in the 700-1000 nm region, which contains half of the NIR solar energy and a quarter of the total solar energy received by North America (Levinson et al., 2005*a*; Levinson et al., 2005*b*). This would indicate that phthalocyanine green's efficacy over phthalocyanine blue is related to an increase in NIR absorbance, and a potentially greater build-up of heat.

In materials science, interfacial heat transfer is characterized by thermal boundary resistance, which causes a temperature discontinuity at the interface of two materials. This is important, as the efficiency of thermal transport across dissimilar interfaces (i.e.-copper phthalocyanine/plant) will determine levels of heat dissipation (Jin, Yadav, K. Sun, H. Sun, Pipe, & Shtein, 2011). Jin et al. (2011) recently discovered a low thermal conductivity between copper phthalocyanine and metal, indicating the presence of a large thermal boundary resistance. Plant leaves naturally have lower thermal conductivity than metals (Dixon & Grace, 1983).

Phthalocyanine blue and phthalocyanine green are both insoluble, taking the appearance of powders. The weak scattering properties of these copper phthalocyanines are due to their small particle size, which is typically 120 nm in diameter (Levinson et al., 2005b). Chlorophyll is also water-insoluble; however chlorophyll *a* and *b* molecules are much smaller at less than 2 nm in diameter (Yu, Yang, & Kim 2010; Cofrancesco, 2000; Strebeyko, 2000).

*Chlorella*, a unicellular green algae, was first recognized through isolation by Beijernick (1890). Among known photosynthetic organisms, it is highest in chlorophyll production, capable of performing photosynthesis at a rate much higher than that of many plant species. Its biomass also contains high concentrations of carotenoid pigments that are capable of providing unique health benefits in humans (Cha, Koo, & Lee, 2008). Since their introduction to the health market during the 1960s, *Chlorella* species have experienced a pronounced growth in production for use as health supplements. One of the most popular species for these applications is *Chlorella vulgaris* (Kanno, 2005). Capable of being cultivated in large scale bioreactors, *Chlorella vulgaris* holds a significant advantage over that of higher plant species, providing a cheap and reliable source for the mass production of beneficial nutrients (Scragg, Illman, Carden, & Shales, 2002).

Ranging from 2 to 10  $\mu\text{m}$  in diameter, *Chlorella vulgaris* cells have a globular shape, and a strengthened cell wall that prevents its adequate digestion and beneficial uptake in humans. For this reason, its cells are fragmented following cultivation, allowing its cell contents, particularly lutein, to have greater bioavailability in humans (Mitsuda, Nishikawa, Higuchi, Nakajima, & Kawai, 1977; Shibata & Hayakawa, 2009). Commercially available forms of *Chlorella vulgaris* thus consist of fragmented cells sold as a powder (Görs, Schumann, Hepperle, & Karsten, 2010).

The absorption spectrum of *Chlorella vulgaris* has light attenuation properties similar to that of chlorinated copper phthalocyanine, with peaks in the NIR region at 600 to 700 nm and the UV-blue light region at 400-500nm (Ley & Mauzerall, 1982; Yun & Park, 2001). It is hypothesized that the foliar application of pulverized *Chlorella vulgaris* cells can improve creeping bentgrass health under suboptimal levels of light exposure.

## References

- Abu-Hamdeh, N. H., & Reeder, R. C. (2000). Soil thermal conductivity: Effects of density, moisture, salt concentration, and organic matter. *Soil Science Society of America Journal*, *64*(4), 1285-1290.
- Allakhverdiev, S. I., & Murata, N. (2004). Environmental stress inhibits the synthesis de novo of proteins involved in the photodamage-repair cycle of photosystem II in *Synechocystis* sp. PCC 6803. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, *1657*(1), 23-32.
- Anderson, J. M., & Andersson, B. (1988). The dynamic photosynthetic membrane and regulation of solar energy conversion. *Trends in Biochemical Sciences*, *13*(9), 351-355.
- Anderson, J. M., Park, Y.-I., & Chow, W. S. (1998). Unifying model for the photoinactivation of Photosystem II in vivo under steady-state photosynthesis. *Photosynthesis Research*, *56*(1), 1-13. doi: 10.1023/a:1005946808488
- Apel, K., & Hirt, H. (2004). REACTIVE OXYGEN SPECIES: Metabolism, Oxidative Stress, and Signal Transduction. *Annual Review of Plant Biology*, *55*(1), 373-399. doi: doi:10.1146/annurev.arplant.55.031903.141701
- Aro, E.-M., Suorsa, M., Rokka, A., Allahverdiyeva, Y., Paakkarinen, V., Saleem, A., . . . Rintamäki, E. (2005). Dynamics of photosystem II: a proteomic approach to thylakoid protein complexes. *Journal of Experimental Botany*, *56*(411), 347-356. doi: 10.1093/jxb/eri041
- Asada, K. (1999). The water-water cycle in chloroplasts: scavenging of active oxygens and dissipation of excess photons. *Annual Review of Plant Biology*, *50*(1), 601-639.
- ASTM, G. 173-03. (2003). *Standard Tables for Reference Solar Spectral Irradiances: Direct Normal and Hemispherical on 37° Tilted Surface*.
- Atwell, B. J., Kriedemann, P. E., & Turnbull, C. G. N. (1999). *Plants in action: adaptation in nature, performance in cultivation*: Macmillan Co of Australia.
- Barber, J. (2008). Crystal Structure of the Oxygen-Evolving Complex of Photosystem II. *Inorganic Chemistry*, *47*(6), 1700-1710. doi: 10.1021/ic701835r
- Beard, J. B. (1973). *Turfgrass: Science and culture*: Prentice Hall Upper Saddle River, NJ.
- Beijerinck, M.W., 1890. Kulturversuche mit Zoochloren, Lichenengonidien und anderen niederen Algen. *Bot. Ztg.* *48*, 725–785.

- Bhagwat, S., Gebhardt, S., Haytowitz, D., Holden, J., & Harnly, J. (2006). USDA Database for the Flavonoid Content of Selected Foods. Release 2. *USDA National Nutrient Database for Standard Reference*.
- Bigger, S., & Delatycki, O. (1989). The effects of pigments on the photostability of polyethylene. *Journal of Materials Science*, 24(6), 1946-1952. doi: 10.1007/bf02385404
- Bohn, T., & Walczyk, T. (2004). Determination of chlorophyll in plant samples by liquid chromatography using zinc-phthalocyanine as an internal standard. *Journal of Chromatography A*, 1024(1-2), 123-128. doi: DOI: 10.1016/j.chroma.2003.10.067
- Bohr, N. (1949). Discussion with Einstein on epistemological problems in atomic physics: University of Copenhagen.
- Bristow, K., & Horton, R. (1996). Modeling the impact of partial surface mulch on soil heat and water flow. *Theoretical and Applied Climatology*, 54(1), 85-98.
- Brønnum-Hansen, K., Jacobsen, F., & Flink, J. (1985). Anthocyanin colourants from elderberry (*Sambucus nigra* L.). 1. Process considerations for production of the liquid extract. *International Journal of Food Science & Technology*, 20(6), 703-711.
- Campbell, G. S., Jungbauer, J. D. J., Bidlake, W. R., & Hungerford, R. D. (1994). Predicting the Effect of Temperature on Soil Thermal Conductivity. *Soil Science*, 158(5), 307-313.
- Carlsen, C., & Stapelfeldt, H. (1997). Light sensitivity of elderberry extract. quantum yields for photodegradation in aqueous solution. *Food Chemistry*, 60(3), 383-387. doi: Doi: 10.1016/s0308-8146(96)00356-1
- Carrow, R. N. (1996). Summer decline of bentgrass greens. *Golf Course Management*, 64(6), 51-56.
- Carrow, R. N. (2004). Surface organic matter in creeping bentgrass greens. *Golf Course Management*, 72(5), 96-101.
- Cha, K. H., Koo, S. Y., & Lee, D.-U. (2008). Antiproliferative Effects of Carotenoids Extracted from *Chlorella ellipsoidea* and *Chlorella vulgaris* on Human Colon Cancer Cells. *Journal of Agricultural and Food Chemistry*, 56(22), 10521-10526. doi: 10.1021/jf802111x

- Chu, C. W., Shrotriya, V., Li, G., & Yang, Y. (2006). Tuning acceptor energy level for efficient charge collection in copper-phthalocyanine-based organic solar cells. *Applied physics letters*, 88, 153504.
- Close, D., & Beadle, C. (2003). The ecophysiology of foliar anthocyanin. *The Botanical Review*, 69(2), 149-161. doi: 10.1663/0006-8101(2003)069[0149:teofa]2.0.co;2
- Cofrancesco, A. J. (2000). *Dyes, Natural*: John Wiley & Sons, Inc.
- Cornic, G., & Ghashghaie, J. (1991). Effect of temperature on net CO<sub>2</sub> assimilation and photosystem II quantum yield of electron transfer of French bean (<i>Phaseolus vulgaris</i> L.) leaves during drought stress. *Planta*, 185(2), 255-260. doi: 10.1007/bf00194068
- Demmig-Adams, B., & Adams III, W. W. (1996). The role of xanthophyll cycle carotenoids in the protection of photosynthesis. *Trends in Plant Science*, 1(1), 21-26.
- DeRose, V. J., Mukerji, I., Latimer, M. J., Yachandra, V. K., Sauer, K., & Klein, M. P. (1994). Comparison of the Manganese Oxygen-Evolving Complex in Photosystem II of Spinach and *Synechococcus* sp. with Multinuclear Manganese Model Compounds by X-ray Absorption Spectroscopy. *Journal of the American Chemical Society*, 116(12), 5239-5249. doi: 10.1021/ja00091a031
- Diffey, B. L. (2002). What is light? *Photodermatology, Photoimmunology & Photomedicine*, 18(2), 68-74.
- Dixon, M., & Grace, J. (1983). Natural convection from leaves at realistic Grashof numbers. *Plant, Cell & Environment*, 6(8), 665-670. doi: 10.1111/1365-3040.ep11589240
- Duysens, L., & Ames, J. (1962). Function and identification of two photochemical systems in photosynthesis. *Biochimica et Biophysica Acta*, 64(2), 243-260.
- Farag, A. A. M. (2007). Optical absorption studies of copper phthalocyanine thin films. *Optics & Laser Technology*, 39(4), 728-732. doi: DOI: 10.1016/j.optlastec.2006.03.011
- Forbes, J. C., & Watson, R. D. (1992). *Plants in agriculture*: Cambridge Univ Pr.
- Fossen, T., Cabrita, L., & Andersen, Ø. M. (1998). Colour and stability of pure anthocyanins influenced by pH including the alkaline region. *Food Chemistry*. ISSN, 0308-8146.

- Gaffen, D. J., & Ross, R. J. (2010). Climatology and trends of US surface humidity and temperature.
- Gates, D. M. (1965). Energy, Plants, and Ecology. *Ecology*, 46(1/2), 1-13.
- Görs, M., Schumann, R., Hepperle, D., & Karsten, U. (2010). Quality analysis of commercial Chlorella products used as dietary supplement in human nutrition. *Journal of Applied Phycology*, 22(3), 265-276.
- Gould, K. (2010). Muriel Wheldale Onslow and the Rediscovery of Anthocyanin Function in Plants. *Recent Advances in Polyphenol Research*, 206.
- Gould, K., McKelvie, J., & Markham, K. (2002). Do anthocyanins function as antioxidants in leaves? Imaging of H<sub>2</sub>O<sub>2</sub> in red and green leaves after mechanical injury. *Plant, Cell & Environment*, 25(10), 1261-1269.
- Gounaris, K., Mannock, D. A., Sen, A., Brain, A. P. R., Williams, W. P., & Quinn, P. J. (1983). Polyunsaturated fatty acyl residues of galactolipids are involved in the control of bilayer/non-bilayer lipid transitions in higher plant chloroplasts. *Biochimica et Biophysica Acta (BBA) - Biomembranes*, 732(1), 229-242. doi: 10.1016/0005-2736(83)90207-9
- Hakala, M., Tuominen, I., Keränen, M., Tyystjärvi, T., & Tyystjärvi, E. (2005). Evidence for the role of the oxygen-evolving manganese complex in photoinhibition of Photosystem II. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 1706(1-2), 68-80. doi: DOI: 10.1016/j.bbabi.2004.09.001
- Harborne, J. B., & Smith, D. M. (1978). Correlations between anthocyanin chemistry and pollination ecology in the polemoniaceae. *Biochemical Systematics and Ecology*, 6(2), 127-130. doi: Doi: 10.1016/0305-1978(78)90038-8
- Hawley, A. J. (1971). Remote Sensing: With Special Reference to Agriculture and Forestry (Vol. 61, pp. 316-318): JSTOR.
- Hernández, I., Alegre, L., Van Breusegem, F., & Munné-Bosch, S. (2009). How relevant are flavonoids as antioxidants in plants? *Trends in Plant Science*, 14(3), 125-132.
- Hipskind, J., Wood, K., & Nicholson, R. (1996). Localized stimulation of anthocyanin accumulation and delineation of pathogen ingress in maize genetically resistant to *Bipolaris maydis* race O. *Physiological and Molecular Plant Pathology*, 49(4), 247-256.

- Hoch, W., Singaas, E., & McCown, B. (2003). Resorption protection. Anthocyanins facilitate nutrient recovery in autumn by shielding leaves from potentially damaging light levels. *Plant physiology*, *133*(3), 1296.
- Hormaetxe, K., Becerril, J. M., Fleck, I., Pintó, M., & García-Plazaola, J. I. (2005). Functional role of red (retro)-carotenoids as passive light filters in the leaves of *Buxus sempervirens* L.: increased protection of photosynthetic tissues? *Journal of Experimental Botany*, *56*(420), 2629-2636. doi: 10.1093/jxb/eri255
- Huang, B. (2002). Getting to the Root of Summer Bentgrass Decline. *USGA Green Section Record*, *40*(4), 21-23.
- Huang, J., Yu, J., Lin, H., & Jiang, Y. (2009). Detailed analysis of bathocuproine layer for organic solar cells based on copper phthalocyanine and  $C_{60}$ . *Journal of Applied Physics*, *105*(7), 073105-073105-073105.
- Hudson, R., & Shane, S. (1994). Organic matter comparison of wettable and nonwetable soils from bentgrass sand greens. *Soil Science Society of America Journal*, *58*(2), 361.
- Hughes, N. M., & Smith, W. K. (2007). Attenuation of incident light in *Galax urceolata* (Diapensiaceae): concerted influence of adaxial and abaxial anthocyanic layers on photoprotection. *Am. J. Bot.*, *94*(5), 784-790. doi: 10.3732/ajb.94.5.784
- Hummel, N. (1993). Rationale for the revisions of the USGA green construction specifications. *USGA Green Section Record*. March/April, 7-21.
- Jin, Y., Yadav, A., Sun, K., Sun, H., Pipe, K. P., & Shtein, M. (2011). Thermal boundary resistance of copper phthalocyanine-metal interface. *Applied physics letters*, *98*(9), 093305-093303. doi: 10.1063/1.3555449
- Kadish, K. M., Smith, K. M., & Guillard, R. (2003). *The Porphyrin Handbook: Applications of phthalocyanines*: Academic Pr.
- Kangatharalingam, N., Pierce, M. L., Bayles, M. B., & Essenberg, M. (2002). Epidermal anthocyanin production as an indicator of bacterial blight resistance in cotton. *Physiological and Molecular Plant Pathology*, *61*(3), 189-195. doi: DOI: 10.1006/pmpp.2002.0434

- Kanno, T. (2005). *Chlorella Vulgaris and Chlorella Vulgaris Extract (CVE): The Powerful Japanese Medicinal Green Algae as a Biological Response Modifier*: Woodland Publishing.
- Karageorgou, P., & Manetas, Y. (2006). The importance of being red when young: anthocyanins and the protection of young leaves of *Quercus coccifera* from insect herbivory and excess light. *Tree Physiology*, 26(5), 613.
- Karan, S., Basak, D., & Mallik, B. (2007). Copper phthalocyanine nanoparticles and nanoflowers. *Chemical Physics Letters*, 434(4-6), 265-270. doi: DOI: 10.1016/j.cplett.2006.12.007
- Karcher, D. E., & Richardson, M. D. (2003). Quantifying Turfgrass Color Using Digital Image Analysis. *Crop Sci.*, 43(3), 943-951. doi: 10.2135/cropsci2003.9430
- Knipling, E. B. (1970). Physical and physiological basis for the reflectance of visible and near-infrared radiation from vegetation. *Remote Sensing of Environment*, 1(3), 155-159. doi: Doi: 10.1016/s0034-4257(70)80021-9
- Kopsell, D. A., Lefsrud, M. G., Sorochan, J. C., & McElroy, J. S. (2010). Pigment Concentrations among Heat-tolerant Turfgrasses. *HortScience*, 45, 650-653.
- Larcher, W. (2003). *Physiological plant ecology: ecophysiology and stress physiology of functional groups*: Springer Verlag.
- Larkindale, J., & Huang, B. (2004). Changes of lipid composition and saturation level in leaves and roots for heat-stressed and heat-acclimated creeping bentgrass (*Agrostis stolonifera*). *Environmental and Experimental Botany*, 51(1), 57-67. doi: Doi: 10.1016/s0098-8472(03)00060-1
- Larkindale, J., & Huang, B. (2004). Changes of lipid composition and saturation level in leaves and roots for heat-stressed and heat-acclimated creeping bentgrass (*Agrostis stolonifera*). *Environmental and Experimental Botany*, 51(1), 57-67. doi: Doi: 10.1016/s0098-8472(03)00060-1
- Larkindale, J., & Knight, M. R. (2002). Protection against Heat Stress-Induced Oxidative Damage in *Arabidopsis* Involves Calcium, Abscisic Acid, Ethylene, and Salicylic Acid. *Plant physiology*, 128(2), 682-695. doi: 10.1104/pp.010320
- Larson, R. A. (1988). The antioxidants of higher plants. *Phytochemistry*, 27(4), 969-978.

- Levinson, R., Berdahl, P., & Akbari, H. (2005a). Solar spectral optical properties of pigments--part I: model for deriving scattering and absorption coefficients from transmittance and reflectance measurements. *Solar Energy Materials and Solar Cells*, 89(4), 319-349.
- Levinson, R., Berdahl, P., & Akbari, H. (2005b). Solar spectral optical properties of pigments--part II: survey of common colorants. *Solar Energy Materials and Solar Cells*, 89(4), 351-389.
- Ley, A. C., & Mauzerall, D. C. (1982). Absolute absorption cross-sections for Photosystem II and the minimum quantum requirement for photosynthesis in *Chlorella vulgaris*. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 680(1), 95-106. doi: 10.1016/0005-2728(82)90320-6
- Liu, X., & Huang, B. (2000). Heat Stress Injury in Relation to Membrane Lipid Peroxidation in Creeping Bentgrass. *Crop Sci.*, 40(2), 503-510. doi: 10.2135/cropsci2000.402503x
- Liu, X., & Huang, B. (2008). Photosynthetic acclimation to high temperatures associated with heat tolerance in creeping bentgrass. *Journal of plant physiology*, 165(18), 1947-1953. doi: 10.1016/j.jplph.2008.05.001
- Loomis, W. E. (1965). Absorption of Radiant Energy by Leaves. *Ecology*, 46(1/2), 14-17.
- Loughrin, J. H., & Kasperbauer, M. J. (2003). Aroma Content of Fresh Basil (*Ocimum basilicum* L.) Leaves Is Affected by Light Reflected from Colored Mulches. *Journal of Agricultural and Food Chemistry*, 51(8), 2272-2276. doi: 10.1021/jf021076c
- Ludwig, C., Strohmaier, R., Petersen, J., Gompf, B., & Eisenmenger, W. (1994). Epitaxy and scanning tunneling microscopy image contrast of copper<math>\text{Cu}</math>phthalocyanine on graphite and  $\text{MoS}_2$ . *Journal of Vacuum Science & Technology B: Microelectronics and Nanometer Structures*, 12(3), 1963-1966.
- McCree, K. J. (1971). The action spectrum, absorptance and quantum yield of photosynthesis in crop plants. *Agricultural Meteorology*, 9, 191-216. doi: 10.1016/0002-1571(71)90022-7
- McDonald, M. S. (2003). *Photobiology of higher plants*: John Wiley & Sons Inc.
- McElroy, J. S., Kopsell, D. A., Sorochan, J. C., & Sams, C. E. (2006). Response of Creeping Bentgrass Carotenoid Composition to High and Low Irradiance. *Crop Sci.*, 46(6), 2606-2612. doi: 10.2135/cropsci2006.02.0119

- Merzlyak, M., Melo, T., & Naqvi, K. (2008). Effect of anthocyanins, carotenoids, and flavonols on chlorophyll fluorescence excitation spectra in apple fruit: signature analysis, assessment, modelling, and relevance to photoprotection. *Journal of Experimental Botany*, 59(2), 349.
- Miller, W., Akbari, H., & Levinson, R. (2004). Special infrared reflective pigments make a dark roof reflect almost like a white roof. *Thermal Performance of the Exterior Envelopes of Buildings, Proceedings of ASHRAE THERM*, Clearwater, FL.
- Mitsuda, H., Nishikawa, Y., Higuchi, M., Nakajima, K., & Kawai, F. (1977). Effect of the breaking of Chlorella cells on the digestibility of Chlorella protein. *J Jpn Soc Food Nutr*, 30, 93-98.
- Moore, J. F. (2004). Revising the USGA's recommendations for a method of putting green construction. *USGA Green Section Record*, 42(3), 1-4.
- Muller, P., Li, X. P., & Niyogi, K. K. (2001). Non-photochemical quenching. A response to excess light energy. *Plant physiology*, 125(4), 1558.
- NCDC (2010, October 21). *U.S. Heat Stress Index*. Retrieved from <http://www.ncdc.noaa.gov/temp-and-precip/heat-stress.html>
- NCDC (2011, April 7). *Time series of number of days average apparent temperature exceeded the 1961 - 90 85th percentile threshold at: KNOXVILLE TN*. Retrieved from [ftp://ftp.ncdc.noaa.gov/pub/data/heatstress/stationdata/KNOXVILLE\\_TN\\_1day\\_ave.ts](ftp://ftp.ncdc.noaa.gov/pub/data/heatstress/stationdata/KNOXVILLE_TN_1day_ave.ts)
- Neil Hunter, C., van Grondelle, R., & Olsen, J. D. (1989). Photosynthetic antenna proteins: 100 ps before photochemistry starts. *Trends in Biochemical Sciences*, 14(2), 72-76. doi: 10.1016/0968-0004(89)90047-9
- Nishio, J. N. (2000). Why are higher plants green? Evolution of the higher plant photosynthetic pigment complement. *Plant, Cell & Environment*, 23(6), 539-548. doi: 10.1046/j.1365-3040.2000.00563.x
- Nishiyama, Y., Yamamoto, H., Allakhverdiev, S. I., Inaba, M., Yokota, A., & Murata, N. (2001). Oxidative stress inhibits the repair of photodamage to the photosynthetic machinery. [10.1093/emboj/20.20.5587]. *EMBO J*, 20(20), 5587-5594.
- NOAA (2010, October 5). *State of the Climate: National Overview for August 2010*. Retrieved from <http://www.ncdc.noaa.gov/sotc/national/2010/8>

- Norton, L. H., Hanrahan, R. K., & Spak, D. R. (2004). Method of improving turfgrass quality: US Patent App. 20,050/181,949.
- Norton, L. H., Hanrahan, R. K., & Spak, D. R. (2005). Method of improving turfgrass quality: US Patent App. 20,060/068,991.
- Norton, L., Hanrahan, R., & Spak, D. (2006). Method of improving grass quality: Google Patents.
- Ohnishi, N., Allakhverdiev, S. I., Takahashi, S., Higashi, S., Watanabe, M., Nishiyama, Y., & Murata, N. (2005). Two-Step Mechanism of Photodamage to Photosystem II: Step 1 Occurs at the Oxygen-Evolving Complex and Step 2 Occurs at the Photochemical Reaction Center†. *Biochemistry*, *44*(23), 8494-8499. doi: 10.1021/bi047518q
- Pallardy, S. G., & Kozlowski, T. T. (2008). *Physiology of woody plants*: Academic Press.
- Planck, M. (1989). *The theory of heat radiation*: Amer Inst of Physics.
- Powles, S. B. (1984). Photoinhibition of Photosynthesis Induced by Visible Light. *Annual Review of Plant Physiology*, *35*(1), 15-44. doi: doi:10.1146/annurev.pp.35.060184.000311
- Prasad, M. N. V. (1997). *Plant ecophysiology*: Wiley.
- Rawson, H., Begg, J., & Woodward, R. (1977). The effect of atmospheric humidity on photosynthesis, transpiration and water use efficiency of leaves of several plant species. *Planta*, *134*(1), 5-10.
- Rice-Evans, C., Miller, N., & Paganga, G. (1997). Antioxidant properties of phenolic compounds. *Trends in Plant Science*, *2*(4), 152-159. doi: Doi: 10.1016/s1360-1385(97)01018-2
- Richerson, D. W. (2006). *Modern ceramic engineering: properties, processing, and use in design*: CRC Press.
- Sarkar, D., Bhowmik, P. C., Young-In-Kwon, & Shetty, K. (2009). Cold Acclimation Responses of Three Cool-season Turfgrasses and the Role of Proline-associated Pentose Phosphate Pathway. *J. Amer. Soc. Hort. Sci.*, *134*(2), 210-220.
- Saron, C., Zulli, F., Giordano, M., & Felisberti, M. I. (2006). Influence of copper-phthalocyanine on the photodegradation of polycarbonate. *Polymer Degradation and Stability*, *91*(12), 3301-3311. doi: DOI: 10.1016/j.polymdegradstab.2006.06.004

- Scragg, A. H., Illman, A. M., Carden, A., & Shales, S. W. (2002). Growth of microalgae with increased calorific values in a tubular bioreactor. *Biomass and Bioenergy*, 23(1), 67-73. doi: 10.1016/s0961-9534(02)00028-4
- Shibata, S., & Hayakawa, K. (2009). Bioavailability of Lutein in Chlorella Powder: A Single Ingestion of Chlorella Powder Raises Serum Lutein Concentrations in Healthy Human Volunteers. *Food science and technology research*, 15(4), 449-452.
- Smith, J. H. C., & French, C. S. (1963). The Major and Accessory Pigments in Photosynthesis. *Annual Review of Plant Physiology*, 14(1), 181-224. doi: doi:10.1146/annurev.pp.14.060163.001145
- Solovchenko, A., & Merzlyak, M. (2008). Screening of visible and UV radiation as a photoprotective mechanism in plants. *Russian Journal of Plant Physiology*, 55(6), 719-737.
- Song, Y., Liu, B., Wang, L., Li, M., & Liu, Y. (2006). Damage to the oxygen-evolving complex by superoxide anion, hydrogen peroxide, and hydroxyl radical in photoinhibition of photosystem II. *Photosynthesis Research*, 90(1), 67-78. doi: 10.1007/s11120-006-9111-7
- Steadman, R. G. (1984). A universal scale of apparent temperature. *Journal of climate and applied meteorology*, 23(12), 1674-1687.
- Steyn, W. J., Wand, S. J. E., Holcroft, D. M., & Jacobs, G. (2002). Anthocyanins in vegetative tissues: a proposed unified function in photoprotection. *New Phytologist*, 155(3), 349-361. doi: 10.1046/j.1469-8137.2002.00482.x
- Strebeyko, P. (2000). Size and Power of Chlorophyll &lt;i>a</i> &lt;i>b</i> Molecule. *Photosynthetica*, 38(1), 159-160. doi: 10.1023/a:1026720729911
- Sun, J., Nishio, J. N., & Vogelmann, T. C. (1996). High-light effects on CO<sub>2</sub> fixation gradients across leaves. *Plant, Cell & Environment*, 19(11), 1261-1271. doi: 10.1111/j.1365-3040.1996.tb00004.x
- Sundby, C., McCaffery, S., & Anderson, J. M. (1993). Turnover of the photosystem II D1 protein in higher plants under photoinhibitory and nonphotoinhibitory irradiance. *Journal of Biological Chemistry*, 268(34), 25476-25482.

- Takahashi, S., & Badger, M. R. (2010). Photoprotection in plants: a new light on photosystem II damage. *Trends in Plant Science*.
- Takahashi, S., & Murata, N. (2005). Interruption of the Calvin cycle inhibits the repair of photosystem II from photodamage. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1708(3), 352-361.
- Takahashi, S., & Murata, N. (2008). How do environmental stresses accelerate photoinhibition? *Trends in Plant Science*, 13(4), 178-182. doi: DOI: 10.1016/j.tplants.2008.01.005
- Takahashi, S., Milward, S. E., Yamori, W., Evans, J. R., Hillier, W., & Badger, M. R. (2010). The solar action spectrum of photosystem II damage. *Plant Physiol.*, pp.110.155747. doi: 10.1104/pp.110.155747
- Tallis, M., Lin, Y., Rogers, A., Zhang, J., Street, N., Miglietta, F., . . . Taylor, G. (2010). The transcriptome of *Populus* in elevated CO<sub>2</sub> reveals increased anthocyanin biosynthesis during delayed autumnal senescence. *New Phytologist*, 186(2), 415-428.
- Tarara, J. M. (2000). Microclimate modification with plastic mulch. *HortScience*, 35(2), 169-180.
- Taulavuori, E., Tahkokorpi, M., Laine, K., & Taulavuori, K. (2010). Drought tolerance of juvenile and mature leaves of a deciduous dwarf shrub *Vaccinium myrtillus* L. in a boreal environment. *Protoplasma*, 241(1), 19-27.
- Thomas, P. G., Dominy, P. J., Vigh, L., Mansourian, A. R., Quinn, P. J., & Williams, W. P. (1986). Increased thermal stability of pigment-protein complexes of pea thylakoids following catalytic hydrogenation of membrane lipids. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 849(1), 131-140. doi: 10.1016/0005-2728(86)90104-0
- Tracton, A. A. (2006). *Coatings materials and surface coatings*: CRC.
- Tripathi, V., Datta, D., Samal, G., Awasthi, A., & Kumar, S. (2008). Role of exciton blocking layers in improving efficiency of copper phthalocyanine based organic solar cells. *Journal of Non-Crystalline Solids*, 354(19-25), 2901-2904.
- Tyystjärvi, E. (2008). Photoinhibition of Photosystem II and photodamage of the oxygen evolving manganese cluster. *Coordination Chemistry Reviews*, 252(3-4), 361-376. doi: DOI: 10.1016/j.ccr.2007.08.021

- van Wijk, K. J., Roobol-Boza, M., Kettunen, R., Andersson, B., & Aros, E. M. (1997). Synthesis and Assembly of the D1 Protein into Photosystem II: Processing of the C-Terminus and Identification of the Initial Assembly Partners and Complexes during Photosystem II Repair†. *Biochemistry*, *36*(20), 6178-6186.
- Vogelman, T. C., Nishio, J. N., & Smith, W. K. (1996). Leaves and light capture: Light propagation and gradients of carbon fixation within leaves. *Trends in Plant Science*, *1*(2), 65-70. doi: Doi: 10.1016/s1360-1385(96)80031-8
- Warnke, S. (2003). Creeping bentgrass (*Agrostis stolonifera* L.). Turfgrass Biology, Genetics, and Breeding, 175-185.
- Wei, Z., Cady, C. W., Brudvig, G. W., & Hou, H. J. M. Photodamage of a Mn(III/IV)-oxo mixed-valence compound and photosystem II: Evidence that a high-valent manganese species is responsible for UV-induced photodamage of the oxygen-evolving complex in photosystem II. *Journal of Photochemistry and photobiology B: Biology, In Press, Corrected Proof*. doi: DOI: 10.1016/j.jphotobiol.2011.01.017
- Wheldale, M. (1916). *The Anthocyanin Pigments of Plants*. Cambridge University Press, Cambridge.
- Williams, C. A., & Grayer, R. J. (2004). Anthocyanins and other flavonoids. *Natural Product Reports*, *21*(4), 539-573.
- Xu, Q., & Huang, B. (2000). Effects of Differential Air and Soil Temperature on Carbohydrate Metabolism in Creeping Bentgrass. *Crop Sci.*, *40*(5), 1368-1374. doi: 10.2135/cropsci2000.4051368x
- Xu, Q., & Huang, B. (2000). Effects of Differential Air and Soil Temperature on Carbohydrate Metabolism in Creeping Bentgrass. *Crop Sci.*, *40*(5), 1368-1374. doi: 10.2135/cropsci2000.4051368x
- Xu, Q., & Huang, B. (2001). Lowering Soil Temperatures Improves Creeping Bentgrass Growth under Heat Stress. *Crop Sci.*, *41*(6), 1878-1883. doi: 10.2135/cropsci2001.1878
- Yeo, A. R., Caporn, S. J. M., & Flowers, T. J. (1985). The Effect of Salinity upon Photosynthesis in Rice (*Oryza sativa* L.): Gas Exchange by Individual Leaves in relation to their Salt Content. *Journal of Experimental Botany*, *36*(8), 1240-1248. doi: 10.1093/jxb/36.8.1240

- Yu, J. W., Yang, R., & Kim, Y. S. (2010). Differential cytoprotective effect of copper-and iron-containing chlorophyllins against oxidative stress-mediated cell death. *Free Radical Research*, 44(6), 655-667.
- Yun, Y. S., & Park, J. M. (2001). Attenuation of monochromatic and polychromatic lights in *Chlorella vulgaris* suspensions. *Applied Microbiology and Biotechnology*, 55(6), 765-770. doi: 10.1007/s002530100639
- Zhang, X., Ervin, E., & Schmidt, R. (2005). The role of leaf pigment and antioxidant levels in UV-B resistance of dark-and light-green Kentucky bluegrass cultivars. *Journal of the American Society for Horticultural Science*, 130(6), 836-841.
- Zuber, H. (1986). Structure of light-harvesting antenna complexes of photosynthetic bacteria, cyanobacteria and red algae. *Trends in Biochemical Sciences*, 11(10), 414-419. doi: 10.1016/0968-0004(86)90175-1