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Mimicking the Binding and Unbinding of Fe3+ With Transferrin Using A Single Biomimetic Nanochannel

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A new type of apo-Tf functionalized nanochannel is developed to achieve the specific and selective binding and unbinding of Fe$^{3+}$ in a multiple and reversible manner via a protein-conformation-gating mechanism. This ion-responsive biomimetic nanochannel provides a simple, robust, and promising model for studying other biological processes.

Biological ion channels that regulate ion transport and ion selectivity through cell membranes are the basis of many physiological processes in a living organism. Due to the complexity of cell membranes and the limited structural information of ion channels, it is a still very challenging task to study ion transport properties of ion channels in vitro. Recently, development of artificial biomimetic nanochannels as a substitution of ionic channels provides promising model systems to replicate, enhance, and better understand the functionality of these biological channels. Unlike most fragile and complex biological ion channels, artificial biomimetic nanochannels offer several advantages for the fundamental study of different complex biological processes: (1) the nanochannels are usually mechanically and chemically robust, with well-defined channel structures; (2) the nanochannels can specifically mimic certain important functions of ionic channels in cell membranes by isolating other uncertain membrane components; and (3) most importantly, the nanochannels can be readily modified with functional molecules at the inner wall to produce stimuli-response nanochannels for biosensors, molecular filtration, and nanofluidic devices. Thus, a comprehensive understanding of ion behaviors (i.e. ion selectivity, rectification, and gating) in synthetic nanochannels increases our fundamental knowledge of physiological processes in biological ionic channels, but also facilitates the development of practical nanofluidic/nanosensor devices for various biomedical applications.

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Fe³⁺ ions do not bind to the COOH-coated Nanochannel-I even on the nanochannel also changed the surface charge and the surface charge distribution of the nanochannel, which in turn control channel conductivity. Parallel experiments were conducted to confirm the charge distribution of the nanochannel, which can decrease the effective pore size, resulting in low conductivity. Additionally, the formation of Fe³⁺–apo-Tf complexes on the nanochannel also changed the surface charge and the surface charge distribution of the nanochannel, which in turn control channel conductivity. Parallel experiments were conducted to confirm the change in the ionic current was observed (Fig. S3, ESI†), indicating that the apo-Tf functionalized nanochannel indeed exhibits a selective binding to Fe³⁺ ions. Other stimuli-responsive nanochannels also displayed similar I–V behaviors.¹⁹,²⁰

Considering the difficulties in characterizing the physicochemical properties of the surface coating inside the nanochannel, we conducted parallel characterizations on the surface properties of the COOH-SAM-coated gold surface and sodium hydroxide etched PET film before and after surface modifications by apo-Tf, corresponding to the respective surface coatings of Nanochannel I and II. First, we used a surface plasmon resonance (SPR) sensor to real-time monitor the covalent attachment of apo-Tf molecules onto a COOH-SAM-coated gold surface. As shown in Fig. S4A (ESI†), carboxyl groups of COOH-SAM coated on SPR chips were first activated by the injection of a freshly prepared aqueous solution of 15 mg EDC and 3 mg NHS per milliliter for 30 min at room temperature. DI water was injected to obtain a stable baseline. A 2 mg mL⁻¹ apo-Tf solution was flowed over spots of the activated COOH-SAM for 1 hour, followed by washing with DI water to remove unbound or loosely bound proteins. A typical SPR sensogram showed a ~20 nm wavelength shift before and after the apo-Tf injection, corresponding to ~340 ng cm⁻² apo-Tf covalently attached onto the COOH-SAM-coated gold substrate.

Surface wettability of the PET films sequentially modified by NaOH (9 mM) solution, NHS–EDC solution and apo-Tf (2 mg mL⁻¹) solution (corresponding to the same surface modification of Nanochannels I and II) was further examined by water contact angle measurements. Visual inspection of the surface wettability clearly showed a visible change of contact angles on four different PET films. The mean water contact angle was 77.3 ± 4.3° on the untreated PET film (Fig. S4-B2, ESI†). Upon incubation of the untreated PET film in a NaOH solution (9 M) for 4 hours, the contact angle decreased to 32.1 ± 2.2° (Fig. S4-B3, ESI†), indicating the formation of a much more hydrophilic COOH-covered surface on the PET film. The NaOH-etched PET film was further treated with a solution of 15 mg EDC and 3 mg NHS per milliliter for 1 hour at room temperature to initiate the EDC–NHS reaction, and the resulting contact angle of NHS–EDC-activated PET film was changed to 43.4 ± 1.2° (Fig. S4-B4, ESI†), indicating the conversion of –COOH groups to NHS ester groups. Finally, when treating the NHS–EDC-activated film with an apo-Tf aqueous solution of 2 mg mL⁻¹ overnight, the contact angle of apo-Tf-modified PET film decreased to 23.2 ± 1.5° (Fig. S4-B5, ESI†), indicating the successful conjugation of apo-Tf onto the PET film. Thus, the changes of water contact angles also reflect the changes of the surface chemical compositions of the modified PET films.

The specific Fe³⁺ responsive property of the apo-Tf functionalized nanochannel was investigated using fluorescence spectroscopy. Upon selective binding of iron ions to apo-Tf, apo-Tf undergoes a conformational change with Trp residues exposed to the environment, which will decrease the fluorescence intensity of apo-Tf, leading to a direct correlation between iron-binding-induced conformational changes and fluorescence intensity. Fig. 2 compares the fluorescence intensity

Fig. 2  Fluorescence intensity of a fixed concentration of apo-Tf (0.05 mg mL⁻¹) in different ionic solutions.
of a fixed concentration of apo-Tf (0.05 mg mL$^{-1}$) in different ionic solutions. When the apo-Tf was dissolved in DI water, NaCl solution (0.75 mM), and PBS buffer containing NaCl and KCl, separately, the fluorescence curves were almost identical and no fluorescence change was observed. The fluorescence independence of solvent conditions suggests that Na$^+$ and K$^+$ ions do not involve binding interactions with apo-Tf and induce the conformation change of apo-Tf, so that Trp residues are well shielded from other residues. Conversely, when dissolving apo-Tf in FeCl$_3$ solutions of different concentrations, fluorescence intensities were progressively decreased as the FeCl$_3$ concentrations increased from 0.075 to 0.75 mM. The most significant decrease in fluorescence occurred in 0.75 mM FeCl$_3$ solution, instead of in 0.075 and 0.15 mM FeCl$_3$ solutions, suggesting more numerous and stronger bindings formed between Fe$^{3+}$ ions and the unfolded apo-Tf, due in part to greater Coulombic repulsion in the higher Fe$^{3+}$ ionic concentrations. More importantly, compared to the unchanged fluorescence in non-Fe$^{3+}$ solutions, the fluorescence decrease in FeCl$_3$ solutions further confirms that apo-Tf indeed selectively binds to Fe$^{3+}$, not Na$^+$ and K$^+$, as evidenced by the decreased fluorescence density.

In living systems, the reversible binding and unbinding of Fe$^{3+}$ to Tf can (re)cyle over 100 times for iron delivery. Although the exact mechanism for such an overwhelming iron binding-unbinding process still remains unclear, it was proposed$^{24}$ that the Fe$^{3+}$-apo-Tf complex can go through a conformational change from a closed to an open state, and the open state is presumed to be the only conformation capable of forming the mixed ligand intermediate, which can further dissociate into the Fe$^{3+}$-ligand complex and apo-Tf, resulting in the removal of Fe$^{3+}$ from Tf. To mimic the reversible binding-unbinding process between Fe$^{3+}$ and Tf using the nanochannel, nitrotri(methylphosphonic acid) (NTP) (Scheme S1, ES$^f$) was used as a ligand to remove Fe$^{3+}$ from Tf via a ligand exchange reaction, presumably due to a high binding affinity of NTP with Fe$^{3+}$. Fig. 3A shows the open-closed switching ability of the nanochannel for ionic conductivity upon alternately binding and unbinding of Fe$^{3+}$ to and from Tf. After treating the Nanochannel II (i.e. apo-Tf functionalized nanochannel) with a FeCl$_3$ solution (0.75 mM) for 1 hour, Fe$^{3+}$ will specifically bind to Tf causing a decrease of ionic current flowing through the nanochannel from 1.3 nA to 0.3 nA at 2 V using PBS as an electrolyte. Upon Fe$^{3+}$ binding to apo-Tf, which changes the Nanochannel-II to the Nanochannel-III, the Nanochannel-III was then treated with an NTP solution (200 mM) for 1 hour to remove Fe$^{3+}$ from Tf, resulting in the ionic current changing back to 1.3 nA. This reversible and switchable Fe$^{3+}$ binding-unbinding process in the nanochannel can be reproduced repeatedly for 5 cycles, reflecting a protein-conformation-gating ionic transport property.

This reversible binding and removal of Fe$^{3+}$ from the Tf was further tested on the apo-Tf functionalized gold chip. Fig. 3B shows three SPR detecting cycles, and each cycle comprises Fe$^{3+}$ binding by FeCl$_3$ injection, DI water washout, Fe$^{3+}$ removal by NTP injection, DI water washout, and surface regeneration. Each cycle always began with an injection of DI water flowing through the apo-Tf-coated chip to establish the stable baseline. In cycle 1, injection of a FeCl$_3$ solution (0.75 mM), followed by DI water washing, led to an SPR wavelength shift of $\sim$ 1 nm, indicating the binding of Fe$^{3+}$ to the apo-Tf. Then, upon injection of an NTP solution (200 mM), a large wavelength shift of 40 nm was observed, indicating that NTP ligands are immobilized onto the apo-Tf. The subsequent DI water wash caused the baseline to return to its original value, indicating that the attached Fe$^{3+}$ ions were completely removed. At this point, the SPR chip is ready to use in the next cycle. In cycles 2 and 3, similar behaviors of Fe$^{3+}$ binding and unbinding were observed, consistent with multiple reversible iron-binding-unbinding cycles in the nanochannel.

In summary, we developed a new iron-responsive nanochannel to mimic a multiple and reversible binding and unbinding process between Fe ions and transferrins. Integration of apo-Tf into the nanochannel demonstrates a protein-conformation-gating mechanism, which governs the selective binding-unbinding of Fe$^{3+}$ in a reproducible and reversible manner. This work was supported by NSF-CAREER Award CBET-0952624 and NSF-CBET-1158447.

Notes and references